

**AMENDMENTS TO THE SPECIFICATION**

Please amend the specification as shown.

Please insert the enclosed sequence listing between page 40 (the last page of the specification) and page 41 (the first page of the claims).

Please amend paragraphs [0043] to [0047] on pages 36-37, as follows:

[0043] OAT1:

784-810

5'-CTGTGCAGCCTATGCACCCAACTATAC-3' **(SEQ ID NO: 1)**

1218-1190 (antisense strand)

5'-CCTTTGCTTAGAGTCAGTTCCTTCTGCAG-3' **(SEQ ID NO: 2)**

[0044] OAT2:

642-672

5'-CCATCAACTACATCATGTTCGTAAGTCACCCG-3' **(SEQ ID NO: 3)**

1105-1076 (antisense strand)

5'-GATATGTCGGAGCTGAGATGTTCCGAACAG-3' **(SEQ ID NO: 4)**

[0045] OAT3:

437-465

5'-GAGACACCATTGTGATAGAGTGGGACTTG-3' **(SEQ ID NO: 5)**

920-889 (antisense strand)

5'-GATAGAACCAGCCAGCGTATGGACTCTGGTAC-3' **(SEQ ID NO: 6)**

[0046] RST1 (mouse homologue of URAT1)

377-405

5'-CATCTTATGCTTATCCGGGACAAGTCCTC-3' **(SEQ ID NO: 7)**

768-739 (antisense strand)

5'-GAGTCTGTTGAAGAGGGTAGAGCAGTCTAC-3' **(SEQ ID NO: 8)**

[0047] RT-PCR reactions were carried out using Ready-To-Go RT-PCR beads (Amersham). First strand cDNA was synthesized using pd(N)<sub>6</sub> at 42°C for 15 minutes. PCR conditions were initial denaturation at 95°C for 5 min, followed by 32 cycles of denaturation at 95°C for 30 sec, annealing at 55°C for 30 sec, and extension at 72°C for 1 min. Samples were stored at 4°C. PCR products were separated by electrophoresis on 1.5% agarose gel, and bands of appropriate molecular weights were excised from the gel, purified using a QIA quick gel extraction kit (QIAGEN), and subcloned into TOPO TA cloning vector (Invitrogen Corporation). The vector was digested with a restriction enzyme EcoRI, and sequenced by dye terminator method using Applied Biosystem Sequencer (ABI3730). The integrity of poly(A)mRNA was evaluated by amplification of GADPH mRNA using the following primer set: sense primer (5'-ACCCCCAATGTATCCGTTGT-3' (SEQ ID NO: 9)) and antisense primer (5'-TACTCCTTGGAGGCCATGTA-3' (SEQ ID NO: 10)). The result is shown in Fig. 11.